

Synthesis and Characterization of Tin Oxide Nanopowder and Its Application to Sensing Different Pathogens

Akhil Chandran M. K.,¹ Branimir Bajac,² Gregor Filipič,³
Željka Cvejić,⁴ Vladimir V. Srdić,² Milan Radovanović,¹
Mitar Simić,^{1,5} Sohail Sarang,¹ and Goran M. Stojanović^{1*}

¹Faculty of Technical Sciences, University of Novi Sad, Trg Dositeja Obradovića 6, 21000 Novi Sad, Serbia

²Faculty of Technology, University of Novi Sad, Bulevar Cara Lazara 1, 21000 Novi Sad, Serbia

³Jozef Stefan Institute, Jamova 39, Ljubljana, SI 1000, Slovenia

⁴Faculty of Science, University of Novi Sad, Trg Dositeja Obradovića 3, 21000 Novi Sad, Serbia

⁵Faculty of Electrical Engineering, University of Banja Luka, Patre 5, 78000 Banja Luka, Bosnia and Herzegovina

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In this paper, we discuss the processing, fabrication, and characterization of tin oxide (SnO₂)-based sensors for the detection of different pathogens. The sensing properties of SnO₂ coatings sintered at three different temperatures (600, 700, and 800 °C) were demonstrated by impedance microbiology. Sensors for the detection of *Candida albicans* and *Pseudomonas aeruginosa* were manufactured in the form of an interdigitated capacitor (IDC) structure. Electrochemical analysis revealed a change in impedance and a shift in self-resonant frequency (SRF) when the sensor was exposed to bacteria or yeast/fungi media. Structural and morphological characterizations of the nanostructured sensing films were carried out by various analytical techniques including X-ray diffraction, Raman spectroscopy, transmission electron microscopy (TEM), and scanning electron microscopy. The obtained results are promising for the fabrication of robust, cost-effective, and nontoxic SnO₂-based sensors for detecting various pathogens.

1. Introduction

Metal oxides (MOs) and nanomaterials are preferred for the development of different sensors owing to their chemical stability and low production cost.^(1,2) Thick- and thin-film technologies have an essential role in the manufacturing of these sensors.^(3,4) Nanostructured metal oxides (NMOs) such as tin, titanium, zinc, iron, and indium oxides have attracted wide interest in the scientific community because they exhibit interesting characteristics such as biocompatibility, nontoxicity, naturally occurring nanostructures, and catalytic properties. In addition, MOs demonstrate enhanced electron transfer properties, thus improving sensing performances.⁽⁵⁾

Tin oxide (SnO₂) is an intrinsic n-type semiconductor with a band gap of around 3.6 eV^(6,7) and a rutile tetragonal crystal structure.⁽⁸⁾ SnO₂ semiconductors have received significant attention in the fields of sensing,^(9,10) electronics, and catalysis.⁽¹¹⁾ The popularity of SnO₂

*Corresponding author: e-mail: sgoran@uns.ac.rs
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films is due to their good optical, electrical, and catalytic properties, as well as their ability to be prepared with various traditional deposition techniques such as sol-gel deposition, slurry deposition, physical vapor deposition, and chemical vapor deposition. Moreover, recent fabrication techniques have enabled the size reduction of SnO₂-based sensors, increasing their sensitivity and reliability. These techniques include the ultrasonochemical,⁽¹²⁾ DC sputtering, and vacuum evaporation methods.⁽¹³⁾ Despite the fact that very promising antibacterial, anticandidal, and cytotoxic activities have been reported,⁽¹²⁾ as well as H₂ gas sensing properties,⁽¹³⁾ there is still strong interest in the development of synthesis and characterization processes for biomedical applications of SnO₂ films. More precisely, a detailed understanding of the sensing mechanism of SnO₂ in specific applications will identify the advantages and disadvantages of different fabrication methods and substrates. Such conclusions can enable the use of SnO₂ films with enhanced properties in future devices. Because of this, new applications of SnO₂-based sensors and their detailed analysis and characterization are very important for researchers in the field.

One of the ways to enhance the properties of SnO₂ is by reducing the crystallite size to the nanometer range.^(14,15) The electron transfer mechanism is the most valuable property of MO semiconductors for sensor, electronic, and optical applications.⁽¹⁶⁾ The crystallite size, the nature of the surface, and the phase composition are the main parameters that determine electron transport through a material.^(17,18) Furthermore, it is important to develop an inexpensive processing method for realizing user-friendly and miniature devices.⁽¹⁹⁾ Thus, many researchers have recently invested effort in the development and fabrication of cost-effective and compact structures with excellent performances and stability to be used as gas sensors.^(20,21) The methods and procedures used for synthesis have a significant role in controlling the particle size^(22,23) to improve the performance of MO semiconductors.⁽²⁴⁾ The influences of the temperature, pressure, and pH of the reaction medium and sintering process have been studied,⁽²⁵⁾ and it has been concluded that they strongly affect the size of nanoparticles. SnO₂ nanostructures have received particular attention in the development of sensors for the monitoring and detection of different pathogen systems owing to their biocompatibility and the efficient sensing.^(26,27) Recently, the detection of foodborne pathogens has been a focus of study owing to increased concerns about food safety and public health,^(28,29) motivating the development of various sensors for this purpose. The conventional procedure for identifying pathogens usually includes procedures such as culturing microbes and isolation from the pathogen medium, which may take several days to obtain a final result.⁽³⁰⁾ To overcome this time delay, numerous methods for the detection of pathogens have been developed to reduce the assay time.^(31,32) One of the techniques for determining pathogens is impedance spectroscopy, and such a method of detection is called impedance microbiology.⁽³³⁾ Stewart attempted to determine a microorganism by impedance measurement.⁽³⁴⁾ After that, several papers described the impedance-based detection of microorganisms and pathogens.^(35,36) However, there is a lack of articles covering the complete process comprising synthesis of the nanostructured material, creation of the sensing film, manufacturing of the sensor, and its comprehensive characterization and validation.

As reported in this paper, we studied the synthesis and characterization of a SnO₂ nanopowder as well as the sensing performance of SnO₂ films sintered at three different

temperatures (600, 700, and 800 °C). We found that sensing films sintered at the lowest temperature (600 °C) had the smallest grains and the lowest self-resonant frequency (SRF). Such characteristics are key factors enabling the use of low-cost measurement devices because of the low value of the upper-frequency limit that is required. For testing purposes, we applied sensing films sintered at 600 °C to the detection of different pathogens (*Candida albicans* and *Pseudomonas aeruginosa*). Our results showed that SnO₂ films exposed to different pathogens exhibit different patterns of impedance changes and SRF shifts, which can be detected by very simple readout electronics, making these films suitable for *in situ* measurements outside the laboratory.

2. Materials and Methods

2.1 Synthesis of SnO₂ nanopowder

A 1 M solution of SnO₂ was prepared by dissolving Sn(II)Cl₂ (from Alfa Aesar, purity 98%) into 90% absolute ethanol with continuous stirring at 60 °C for about 1 h. The obtained solution was allowed to cool, and the precipitation of SnO₂ was started by the dropwise addition of NaOH solution (0.01 M). NaOH solution was added with continuous stirring until the pH reached 10. The product was collected in a centrifuge flask and washed with water and ethanol several times to remove undesired products from the SnO₂ nanopowder. The resulting product was dried overnight in a drier at 70 °C. The dried powder was collected in a mortar and ground to destroy the formed agglomerations. The powder product was treated at 500 °C in a tube furnace and a pale yellow fine powder of SnO₂ was obtained after calcination (Fig. 1).

2.2 Fabrication procedure of SnO₂-based sensors

The sensor samples were fabricated by screen-printing technology using a previously prepared paste. The paste was obtained by grinding the calcined SnO₂ nanopowder using a

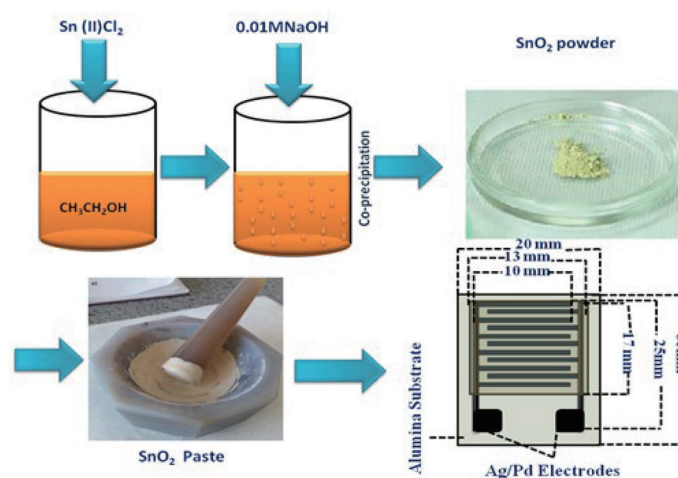


Fig. 1. (Color online) Steps in manufacturing SnO₂ sensing layer on IDC on Al₂O₃ substrate.

planetary ball mill in an isopropanol solvent, and three cycles of grinding were performed with a 1:3 mass ratio of the powder and the grinding ball. The ground powder was then evaporated overnight at 80 °C. The MO paste was made by gently mixing the SnO₂ nanopowder with a binder solution comprising ethyl cellulose and terpineol (1:9 ratio). In the first step of the fabrication, an interdigitated capacitor (IDC) electrode system was formed using a screen-printed Ag/Pd metal layer on a previously cleaned alumina (Al₂O₃) substrate. The IDC structure was composed of eight fingers on one side and eight fingers on the other side, representing the electrodes of a planar capacitor. The width of the conductive lines was 0.8 mm and the space between them was 0.6 mm. This IDC layout and its dimensions were selected to obtain SRFs in the range up to 100 MHz, which is in the range of the HP-4194A commercial impedance analyzer as well as handheld readout electronic devices. The fabrication was completed with the deposition of the SnO₂ paste by a screen printer (Fig. 1). The prepared samples were sintered at 600, 700, and 800 °C and are denoted as SnO₂-600, SnO₂-700, and SnO₂-800, respectively.

2.3 Microbial culture preparation

C. albicans and *P. aeruginosa* pathogen samples were used as bulky media to analyze the sensing properties of SnO₂ coatings, using colonies of *P. aeruginosa* cultured for 24 h on blood agar (HiMedia, India) and colonies of *C. albicans* cultured on Sabouraud dextrose agar (HiMedia, India). In sterile tubes, suspensions with a density of 0.5 MCF (McFarland) were created in 4.5 ml of physiological saline in accordance with the European Committee on Antimicrobial Susceptibility Testing (EUCAST) standard.

2.4 Characterization techniques

The structure of the calcined SnO₂ nanoparticles was analyzed by transmission electron microscopy (TEM) with a JEOL JEM 2100 electron microscope and an acceleration voltage of 200 kV. The surface morphology of the sintered SnO₂ film was determined by scanning electron microscopy (SEM; JEOL JSM6460LV). The phase composition was determined and the structure was characterized by X-ray diffraction (XRD; Rigaku, MiniFlex 600) as well as Raman spectroscopy (Thermo Fisher DXR Raman microscope). The crystallite size was estimated from XRD patterns using the Scherrer equation. The impedance spectroscopic and sensing performance analyses of SnO₂ films towards different pathogen media were conducted using an HP-4194A impedance analyzer (Fig. 2).

3. Results and Discussion

3.1 Characterization of SnO₂ nanopowder

High-resolution TEM (HRTEM) and selected-area electron diffraction (SAED) images of the SnO₂ powder are presented in Fig. 3. It can be seen that the calcined SnO₂ powder is agglomerated and that the observed particles are single-crystalline with a tetragonal rutile

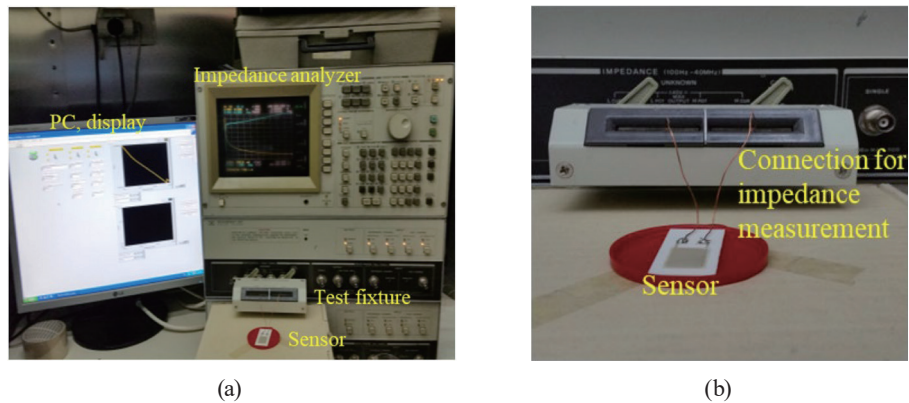


Fig. 2. (Color online) (a) Experimental setup for electrical impedance spectroscopy characterization of the fabricated sensor, and (b) connection of the sensor and the instrument for input impedance measurement.

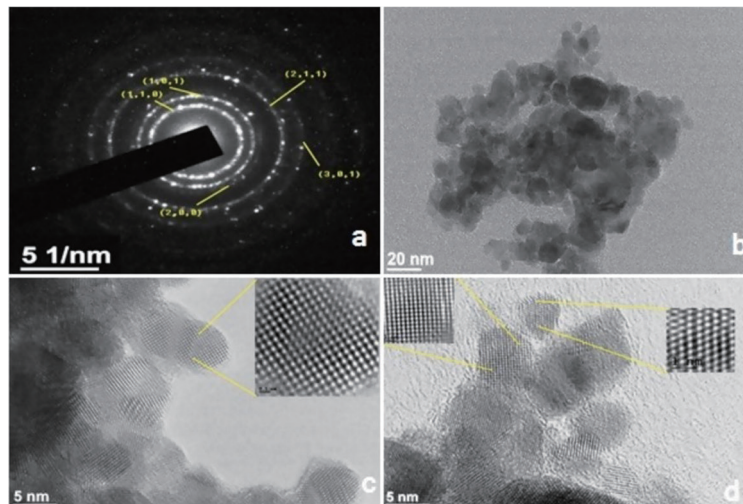


Fig. 3. (Color online) (a) SAED image of SnO₂ powder, (b) TEM image of SnO₂ powder, (c) HRTEM image of SnO₂ powder, and (d) HRTEM image of SnO₂ powder.

structure and nanometer size (less than 25 nm on average). The particle size is very similar to that previously reported,⁽¹²⁾ where the size of the SnO₂ nanoparticles was about 5–30 nm. Moreover, in Ref. 13, the nanostructures had diameters on the order of 10–100 nm and lengths on the order of 10 μm , depending on the type of substrate.

XRD measurements were performed to identify and confirm the crystalline phase of the SnO₂ powder. The XRD spectrum of the calcined SnO₂ powder is shown in Fig. 4(a), from which it can be seen that all the peaks are characteristic peaks of SnO₂ powder without any secondary phase formation. The observed XRD peaks are ascribed to different planes, such as (110), (101), (200), and (211), indicating the formation of the rutile tetragonal structure. In previous

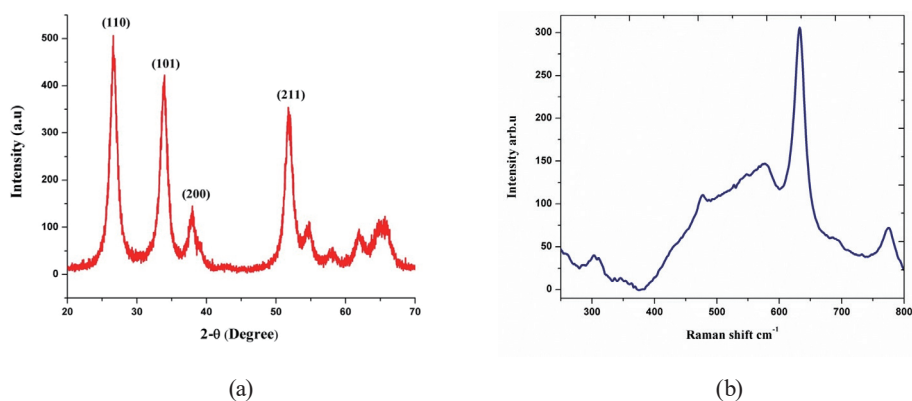


Fig. 4. (Color online) (a) XRD spectrum and (b) Raman spectrum of SnO₂ powder.

studies, strong diffraction peaks ascribed to these planes confirmed the high crystallinity of structures fabricated by the ultrasonochemical method,⁽¹²⁾ as well as the DC sputtering and vacuum evaporation methods.⁽¹³⁾

The formation of SnO₂ nanorods with a tetragonal rutile structure was also indicated by the Raman spectrum. SnO₂ with the rutile structure belongs to the space group P42/mnm, 48–50 with Sn and O atoms in the 2a and 4f positions, respectively. Figure 4(b) depicts the Raman spectrum of the SnO₂ nanorods in the wavenumber range of 300–850 cm⁻¹. The Raman spectrum of the SnO₂ nanorods exhibits the conventional vibration modes at 473, 498, 629, 689, and 770 cm⁻¹.

3.2 Structural characterization of SnO₂ sensing layer

The surface morphology and cross sections of the screen-printed SnO₂ films were studied by SEM, and micrographs are presented in Fig. 5. Figures 5(a) and 5(b) show cross-sectional views of the SnO₂ sensing layer, IDC electrodes, and Al₂O₃ substrate. From the SEM micrograph in Fig. 5(a), the SnO₂ sensing layer of 7.6 μm thickness can be seen, which is clearly distinguishable from the electrode layer with a thickness of 9.5 μm on the Al₂O₃ substrate. Moreover, from the SEM images, it can be seen that each layer is attached without voids to the adjacent layer(s), which explains the good adhesion of the SnO₂ film and IDC electrode system on the Al₂O₃ substrate. Additionally, Fig. 5(c) shows the surface morphology of the SnO₂ sensing coating, where a uniform structure can be observed. A top view of the interdigitated electrode structure can be seen in Fig. 5(d). XRD patterns of the SnO₂-600, SnO₂-700, and SnO₂-800 samples are presented in Fig. 6. All the XRD peaks of the rutile tetragonal SnO₂ structure can be observed. With increasing sintering temperature, the peak intensities increase and the full width at half maximum (FWHM) values decrease.⁽³⁷⁾ These behaviors strongly indicate the enhanced crystallinity and increased grain size in SnO₂.

The average crystallite sizes (D_p) of the sintered SnO₂-600, SnO₂-700, and SnO₂-800 samples were estimated with Scherrer's equation [Eq. (1)] using the FWHM as follows:

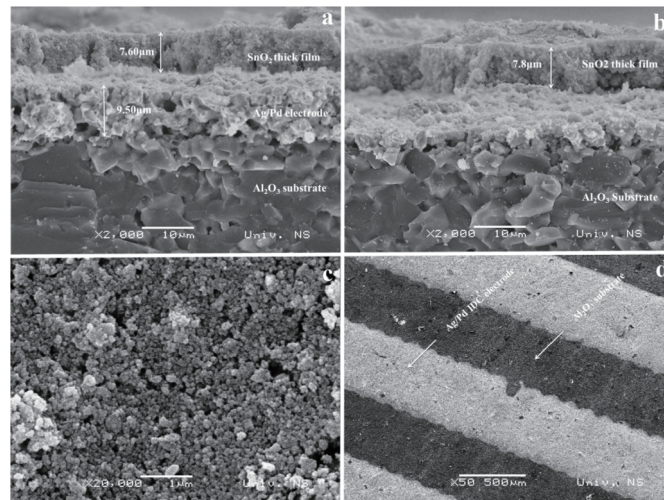


Fig. 5. SEM images of the sensor structure: (a, b) cross-sectional images, (c) surface of SnO₂ layer, and (d) surface of IDC electrodes (fingers).

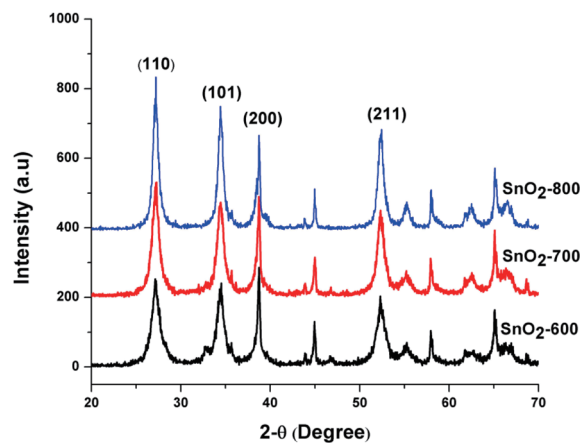


Fig. 6. (Color online) XRD spectra of SnO₂ films (SnO₂-600, SnO₂-700, and SnO₂-800, sintered at 600, 700, and 800 °C, respectively).

$$D_p = \frac{K\lambda}{c \cos \theta \times \beta / 2}, \quad (1)$$

where β is the peak broadening (i.e., FWHM) in radians, λ is the XRD wavelength (0.15405 nm), K is a constant ($K = 0.94$), and θ is the Bragg angle.

The average crystallite sizes of the samples sintered at 600, 700, and 800 °C were 16, 18, and 25 nm, respectively (Table 1), confirming the expected increase in grain size with increasing sintering temperature. The same conclusion has already been reported by Pawar *et al.*⁽³⁸⁾ for nanometer crystallite sizes. The nanometer particle size will enhance the sensitivity of the SnO₂ active layer, as already reported.⁽³⁷⁾

Table 1

Average crystallite size of SnO₂ layer calculated using Scherrer's equation.

Sample	Sintering temperature (°C)	FWHM (°)	Peak position (°)	Crystallite size (nm)
SnO ₂ -600	600	0.525	27.1	16
SnO ₂ -700	700	0.465	27.2	18
SnO ₂ -800	800	0.330	27.1	25

Raman spectra of the sintered SnO₂ films are shown in Fig. 7. From the spectra, the formation of the SnO₂ phase and all major characteristic bands of SnO₂ can be confirmed. The Raman shifts observed at 632 and 472 cm⁻¹ represent the A_{1g} and E_g modes of vibration, respectively, whereas a Raman mode of vibration was observed at 572 cm⁻¹. The Raman shift at 572 cm⁻¹ occurs in SnO₂ with a small grain size. This reveals that the crystallite size of the SnO₂ sensing layer is in the nanometer range.^(39,40) These Raman active modes are the main characteristic of the SnO₂ rutile structure. These spectra confirm that the SnO₂ films were formed in the metal phase.

3.3 Impedance spectroscopic analysis of the proposed sensors

The validation of sensor performances by impedance spectroscopic analysis was performed in two parts as follows:

- (1) Impedance and SRF measurements were performed on the sensors with SnO₂ films sintered at three different temperatures (600, 700, and 800 °C) when they were immersed in a solution of *C. albicans*. The aim of this experiment was to determine the influence of the sintering temperature on the sensor responses (impedance change and SRF) to the same pathogen. It was expected that the grain size, surface roughness, and surface area to volume ratio of the nanostructure would influence the sensing performances, providing valuable information regarding which sintering temperature provides the best sensing performances and enabling the development of low-cost impedance measurement devices for *in situ* applications.
- (2) Impedance and SRF measurements were performed on the sensors with SnO₂ films sintered at the same temperature when they were immersed in solutions with different pathogens (*P. aeruginosa* and *C. albicans*). It was expected that the results would provide clues on how SnO₂ sensing films sintered at the same temperature responded to different pathogens. The presence of pathogens was expected to influence the permittivity/conductivity of the solution and sensing film, producing changes in the impedance and capacitance of the IDC sensor and consequently enabling the identification of specific pathogens through the measured SRF.

As mentioned above, impedance and SRF measurements were performed with an HP-4194A impedance analyzer. The impedance was measured at 60, 120, 180, and 240 kHz at room temperature. The obtained results are provided and discussed in the following subsections.

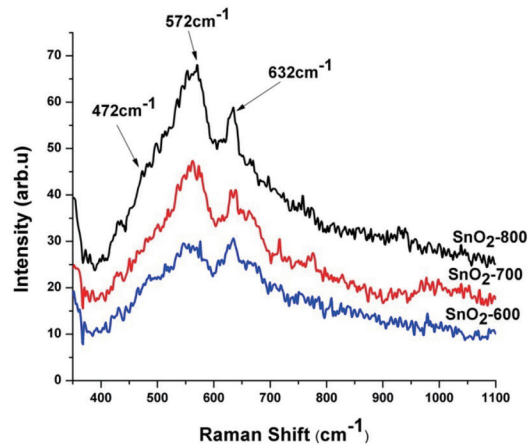


Fig. 7. (Color online) Raman spectra of SnO₂ films sintered at 600, 700, and 800 °C.

3.3.1 Influence of sintering temperature on sensor response to *C. albicans*

The obtained impedance magnitudes and phase angles of the SnO₂ films sintered at three sintering temperatures (600, 700, and 800 °C) when they were exposed to a solution with *C. albicans* are shown in Tables 2 and 3, respectively. The sensor impedance magnitude increased with the sintering temperature at all test frequencies, whereas the impedance phase angle mostly decreased.

Figure 8 presents the capacitance as a function of frequency for the three sensor samples exposed to *C. albicans*. It can be concluded that the SRF increases with increasing sintering temperature.

The higher impedance at the higher sintering temperature indicates that the IDC sensor has lower capacitance and consequently a higher SRF, which means that Fig. 8 supports the values in Table 2. However, because in this experiment the solution was not changed during the measurement, relative changes in impedance should be linked to the influence of the grain size, surface roughness, and surface area to volume ratio of the nanostructure on the sensing performance.

Table 1 shows that the average grain size of the SnO₂ films increased with increasing sintering temperature. It has already been reported that increasing the grain size reduces the porosity of MO films, because the size of grain boundaries decreases as very small pores disappear through diffusion kinetics, whereas the porosity of samples with a smaller grain size is mostly intergranular.⁽⁴¹⁾ Moreover, in Ref. 42, it was shown that the MO structure is homogeneous for a larger grain size and that a more porous structure has a higher capacitance. This means that the sensing film sintered at the lowest temperature (600 °C) in our experiment should exhibit the highest capacitance, and consequently, the lowest SRF, and the sensing film sintered at the highest temperature (800 °C) should exhibit the lowest capacitance and the highest SRF. We obtained such results, as shown in Fig. 8. Moreover, the results in Table 2 are

Table 2

Measured impedance magnitudes Z (Ω) of the sensors sintered at different temperatures.

Sintering temperature ($^{\circ}\text{C}$)	Z (Ω) at 60 kHz	Z (Ω) at 120 kHz	Z (Ω) at 180 kHz	Z (Ω) at 240 kHz
600	11.852	11.719	11.663	11.632
700	13.327	13.160	13.090	13.050
800	16.998	16.823	16.749	16.712

Table 3

Measured impedance phase angles ϕ (deg) of the sensors sintered at different temperatures.

Sintering temperature ($^{\circ}\text{C}$)	ϕ (deg) at 60 kHz	ϕ (deg) at 120 kHz	ϕ (deg) at 180 kHz	ϕ (deg) at 240 kHz
600	-1.938	-0.938	-0.332	0.161
700	-1.935	-1.045	-0.477	-0.016
800	-2.086	-1.096	-0.541	-0.131

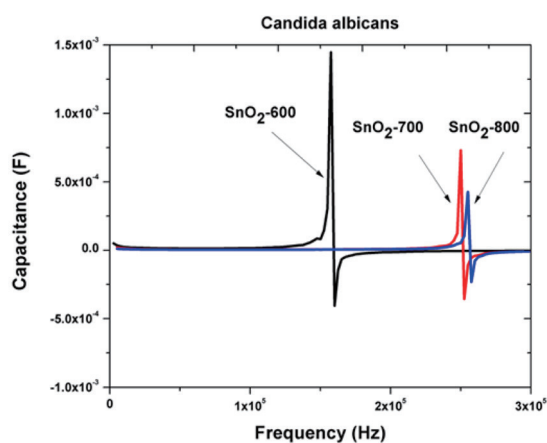


Fig. 8. (Color online) Capacitance as a function of frequency for SnO_2 -600, SnO_2 -700, and SnO_2 -800 in *C. albicans* medium.

in agreement with those in Fig. 8 as there is an increase in impedance with increasing sintering temperature, which means that the sensing film sintered at the lowest temperature had the highest capacitance.

From Fig. 8, it can be concluded that the lowest sintering temperature corresponds to the lowest SRF value. The advantage of a lower SRF is the reduced complexity of the required measurement device, and therefore its cost, as it requires a lower upper-frequency limit, which is very important for *in situ* measurements outside the laboratory. For this reason, we next analyzed the response of SnO_2 -600 films to exposure to different pathogens.

3.3.2 Influence of pathogen medium (*C. albicans* or *P. aeruginosa*) on response of SnO_2 sensor sintered at 600°C

The obtained results for the impedance magnitude and phase angle of sensors sintered at 600°C when they were immersed in solutions with *C. albicans* and *P. aeruginosa* are shown

in Table 4. The sensors had lower impedance magnitudes when they were immersed in a solution with *P. aeruginosa*. However, they exhibited higher phase angles in a solution with *P. aeruginosa*.

Additionally, SRFs were measured when the sensors were placed in a solution with *C. albicans* or *P. aeruginosa*. The obtained SRFs were 157 and 172 kHz for *C. albicans* and *P. aeruginosa*, respectively. Figure 9 shows capacitance as a function of frequency for SnO₂-600 films exposed to *C. albicans* and *P. aeruginosa*, as well as the positions of resonance peaks and the SRF.

Agreement between the results in Table 4 and Fig. 9 can be observed. When the SnO₂-600 sensor was placed in a solution with *C. albicans*, it had higher impedance magnitude at the same frequency than that when the sensor was placed in a solution with *P. aeruginosa*, which means that the IDC sensor had higher capacitance, and consequently lower SRF, when immersed in the solution with *C. albicans*.

This behavior can be attributed to the lower dielectric constant of bacteria than that of yeasts/fungi.⁽⁴³⁾ The analyzed pathogens have different intrinsic structures and characteristics: *C. albicans* is a pathogenic yeast, whereas *P. aeruginosa* is a bacteria. The dielectric properties of yeasts/fungi and bacteria should be different because they have different cellular structures. Yeasts consist of various types of polysaccharides, which produce relatively thick cell walls.⁽⁴³⁾

Table 4

Impedance magnitude and phase angle of sensors sintered at 600 °C immersed in solutions with *C. albicans* and *P. aeruginosa*.

Frequency (kHz)	<i>C. albicans</i>		<i>P. aeruginosa</i>	
	Z (Ω)	φ (deg)	Z (Ω)	φ (deg)
60	11.852	-1.938	7.685	-1.767
120	11.719	-0.938	7.599	-0.615
180	11.663	-0.332	7.563	0.187
240	11.632	0.161	7.544	0.850

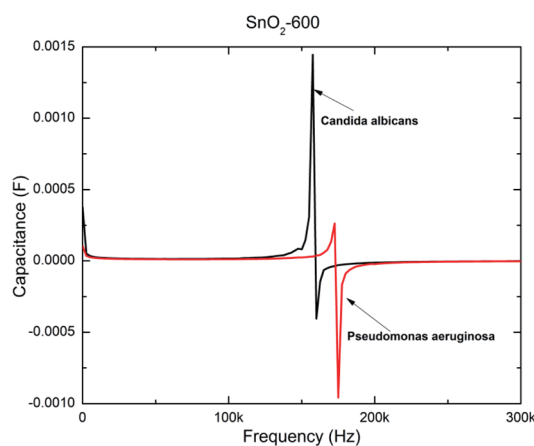


Fig. 9. (Color online) Capacitance as a function of frequency for SnO₂-600 films exposed to *C. albicans* and *P. aeruginosa*.

However, bacterial cell walls are relatively thin (mostly consisting of peptidoglycan). Moreover, yeasts are mainly composed of β -glucan (with long branches and a large molecular weight), while bacteria primarily consist of water. It has been reported that yeasts have a higher dielectric constant than bacteria, which is most probably due to the large polarizability of the branch structure of long-branched β -glucan.⁽⁴¹⁾ These results were also confirmed in an aqueous environment. Additionally, the results from Ref. 41 support our findings presented in Table 4 and Fig. 9.

The results presented in this article are of an initial investigation of the use of SnO₂-based IDC sensors for selective pathogen detection (*P. aeruginosa* and *C. albicans*). We have shown that the different pathogens have different electrical responses, which can be expected from the theoretical analysis of the pathogen structure. For example, yeasts have a higher dielectric constant than bacteria, which led us to expect a higher capacitance and lower SRF for the case of *C. albicans*. Experimental results confirmed this expectation. Therefore, by performing impedance and SRF measurements on a larger number of different pathogens, it will be possible to recognize the unique pattern of changes in the electrical parameters corresponding to specific pathogens. The experimental and processed data in the proposed approach require very small computational resources for their storage, allowing larger datasets and increasing the reliability and accuracy of identification and selective detection.

3.3.3 Comparison of proposed method with other methods for detecting and identifying pathogenic bacteria

The results presented in Sect. 3.3.2 demonstrate an initial step towards the further development of a system for the reliable detection and identification of pathogenic bacteria in real-life applications. Because of this, it is very useful to also give a short overview of phenotypic and molecular diagnostic methods for the detection and identification of pathogenic bacteria, and to compare them with the IDC-sensor-based identification technique.

Phenotypic methods explore biochemical pathways, which are unique for specific bacteria. This approach enables bacterial identification, even though many proteins (and also enzymes) are common to different bacterial species. Phenotypic methods include biochemical testing, the chromogenic media approach, and matrix-assisted laser desorption/ionization–time-of-flight mass spectrometry (MALDI-TOF MS).⁽⁴⁴⁾ During the detection of specific pathogens in biochemical testing, it is necessary to use more specific culture media than general-purpose agar-based media (commonly used in clinical microbiology laboratories). An example of this approach is a differential media target, which combines systems for monitoring nutrition incorporation and indicators for monitoring digested nutrients. Bacteria can be identified with modern automated systems in just a few hours.⁽⁴⁴⁾ In the chromogenic media approach, small amounts of the sample to be analyzed are placed on chromogenic substrates, which are then hydrolyzed. After such treatment, if enzymes of interest are present, the sample will develop a specific color. However, in practice, this approach produces results that require further confirmation by MALDI-TOF MS. In addition, 16 to 48 h is usually required to perform testing.⁽⁴²⁾ MALDI-TOF MS is an ionization technique where ions are created from molecules by means

of a matrix that absorbs laser energy. Identification is based on the time taken by ions to reach the detector, which is dependent on the charge of the ions and characteristic masses. Specialized software (an online database) is then used to compare the obtained mass spectra with defined profiles for specific bacteria. MALDI-TOF MS is suitable for applications where a large number of tests are required every day.⁽⁴²⁾

Hybridization-based detection, amplification methods, and DNA microarrays are the most popular molecular diagnostic methods for bacterial detection. Hybridization-based detection is a very fast and reliable approach for detecting specific bacteria. A detection time of less than 2.5 h and accuracy of 96.5% have been reported.⁽⁴⁴⁾ This approach uses fluorescent dyes with synthetic DNA fragments as probes. The presence of a specific species is determined from the fluorescent signal. Amplification methods are based on the isolation, amplification, and quantification of a short DNA sequence that corresponds to a specific bacterium. These methods usually require between 5 and 24 h for the complete process. DNA microarrays are becoming increasingly popular as modern computer systems allow access to publicly available large-scale whole genome sequencing data. Because of this, genes and combinations of genes of specific pathogens can be determined.

It is evident that, despite some techniques providing promising results, there is still a growing need for rapid, cost-efficient, and reliable systems for bacteria detection and identification. The IDC-sensor-based approach presented in this paper does not require expensive equipment for data processing as the sensing mechanism is based on impedance and SRF measurements. Moreover, the fabrication of IDC sensors is straightforward and economical. In addition, IDC sensors have small dimensions, which make them very suitable for portable systems and measurements outside the laboratory.

4. Conclusions

In this paper, we presented a SnO₂ powder with nanosize particles synthesized by a coprecipitation method. TEM analyses confirmed the presence of the rutile tetragonal structure and a nanometer crystallite size (< 25 nm). Interdigitated capacitive SnO₂ structures were fabricated from this nanopowder using screen-printing technology. Samples were sintered at 600, 700, and 800 °C on an Al₂O₃ substrate. From the calculation using Scherrer's equation, it was confirmed that the grain size of the SnO₂ films was in the range of 16–25 nm. Using impedance spectroscopy, the response of SnO₂ films to *C. albicans* and *P. aeruginosa* was monitored. Impedance spectroscopic studies of the response in a *C. albicans* medium revealed that the sensitivity of the film tended to decrease with increasing sintering temperature and crystallite size. Owing to the lowest crystallite size and highest density of active sites on the surface, the SnO₂-600 film showed the highest sensitivity to the pathogen media. All tested samples showed a characteristic SRF shift that depended on the sensor structure and the exposed pathogen medium.

The main contributions of this article can be summarized as follows: (1) analysis of the effect of the sintering temperature on the sensing characteristics of SnO₂ films, and (2) analysis and explanation of the sensing mechanism of SnO₂ films sintered at the same temperature but

exposed to different pathogen media. The future direction of this study will be oriented towards the development of similar sensor structures on flexible substrates to implement them in real-life applications.

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